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## Enantioselective Synthesis of 7-Cycloocten-1,3,5,6-tetraol Derivatives by Enzymatic Asymmetrization

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**Abstract:** meso-Diol 4, derived from 1,5-cyclooctadiene, in the presence of *Pseudomonas cepacia* lipase in isopropenyl acetate, afforded enantiopure 5, an attractive intermediate for the synthesis of sugars and related compounds.

We are engaged in a program directed toward development of chemoenzymatic syntheses of polyhydroxylated natural products and related bioactive compounds.<sup>1</sup> Using enzymatic asymmetrization<sup>2</sup> and resolution<sup>3</sup> methodologies we have transformed simple hydrocarbons (*e.g.*, cyclopentadiene, benzene or cycloheptatriene) into enantiopure intermediates, which were further elaborated<sup>4</sup> into various targets including sugars and related molecules (Scheme 1).<sup>5</sup>



1,5-Cyclooctadiene, a dimer of 1,3-butadiene and a readily available, inexpensive substance, appeared to be a logical substrate for progression of this strategy. We envisioned that the eight-membered ring intermediate 5 could be a useful starting material for the synthesis of higher sugars and other polyhydroxylated bioactive targets.

Monobromination of cyclooctadiene followed by *cis*-dihydroxylation gave acetonide 1 after acidcatalysed acetalization.<sup>6</sup> Bis-elimination of HBr gave diene 2 (Scheme 2). Examination of molecular models (Figure 1) lead to the prediction that singlet oxygen addition to 2 from the face *syn* to the existing acetonide would be favored. The single peroxide diastereomer 3 formed on exposure of 2 to singlet oxygen<sup>7</sup> had the all *syn* stereochemistry of oxygen substituents in accord with our prediction (vide infra).

Zinc and acetic acid reduction of the peroxide 3 led to diol 4, which was treated with lipase from *Pseudomonas cepacia* (Amano P-30) in isopropenyl acetate at 50 °C for 4 days.<sup>8</sup> Collection of the the enzyme by filtration and evaporation of solvent gave monoacetate 5 in 90% yield. Monacetate 5 was found to be >98% enantiopure by converting alcohol 5 into enone 6 which was analyzed using chiral HPLC.<sup>9</sup> The absolute and relative stereochemistry of 5 was established by chemical correlation to 3-deoxy-D-*ribo*-hexitol pentaacetate (9) (Scheme 3).



4 5 6 <sup>a</sup>Reagents and conditions: (a) Br<sub>2</sub>, CHCl<sub>3</sub>, - 78 °C; (b) OsO<sub>4</sub>-cat, NMO, THF, rt; (c) 2,2dimethoxypropane, p-TsOH-cat, rt; (d) DBU, toluene, reflux: (e) O<sub>2</sub>, TPP, hu, 10 °C, CHCl<sub>3</sub>:MeOH -3:1; (f) Zn(activated), HOAc, EtOAc; (g) isopropenyl acetate, *Pseudomonas cepacia* lipase (Amano P-30) 2 wt. equiv, 50 °C, 4 d



Figure 1. Transannular hydrogens shield approach of  ${}^{1}O_{2}$  anti to acetonide of 2 (mimimized Chem3D<sup>TM</sup> structure shown).

Silylation of the free hydroxy of 5 with TBDPSCl followed by removal of the acetate with KOH/MeOH and PDC oxidation gave enone 7 in 50% yield. Enone 7 was quantitatively converted to the *t*-butyldimethylsilyloxy diene 8. Ozonolysis of 8 followed in sequence by reductive work-up (dimethyl sulfide), sodium borohydride reduction of the dialdehyde, acidic (1N HCl) removal of protective groups and peracetylation gave (+)-3-deoxy-*ribo*-hexitol pentatacetate (9). The spectral data for this compound was identical to that previously reported.<sup>10</sup> The production of diastereomer 9 established the relative stereochemistry of the singlet oxygen addition product 3 and polyols 4 and 5 derivatives as having all syn oxygens.

The absolute stereochemistry was established by conversion of the prostaglandin presursor  $10^{11}$  of known absolute stereochemistry to pentaacetate 9.  $\alpha$ -lodination of enone  $10,^{4a}$  followed by Luche reduction, <sup>12</sup> afforded alcohol 11 as a major product (*syn* : *anti*; 8:1).<sup>5c</sup> One carbon homologation with carbon monoxide<sup>5c,13</sup> followed by reduction and selective silylation of the primary alcohol gave cyclopentenol 12. Ozonolysis of 12 followed by reductive work-up led to a keto-aldehyde, which was immediately subjected to *anti*-selective reduction with sodium triacetoxyborohydride (*anti* : *syn*; >20:1).<sup>14</sup> Acidic removal of the silyl groups and peracetylation again afforded (+)-pentaacetate 9, establishing the absolute confirguration of compounds 5 - 9 as shown.



<sup>a</sup>Reagents and conditions: (a) TBDPSCl, imidazole, DMF, RT; (b) KOH, MeOH; (c) PDC, CH<sub>2</sub>Cl<sub>2</sub>, molecular sieves 4 Å; (d) TBSOTf, Et<sub>3</sub>N, Et<sub>2</sub>O, RT: (e) O<sub>3</sub>, MeOH, -78 °C then DMS, 20 °C; (f) NaBH<sub>4</sub>, MeOH; (g) 1N HCl, MeOH, rt; (h) Ac<sub>2</sub>O, pyridine, DMAP, rt; (i) I<sub>2</sub>, (1.8 eq.), pyridine/CCl<sub>4</sub>; (j) NaBH<sub>4</sub>, CeCl<sub>3</sub>, MeOH, -78 °C; (k) CO (1 atm.), Bu<sub>3</sub>SnH, Pd(Ph<sub>3</sub>P)<sub>4</sub> 5 mol%, THF; then NaBH<sub>4</sub>, CeCl<sub>3</sub>·7H<sub>2</sub>O, MeOH, -78 °C; (l) TBSCl, Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, RT; (m) NaBH(OAc)<sub>3</sub>, AcOH, CH<sub>2</sub>Cl<sub>2</sub>

The absolute stereochemistry at the acetoxy-substituted stereogenic center of 5 is consistent with that predicted by empirical rules for *Pseudomonas cepacia* lipase catalysis at such secondary centers.<sup>2d,15</sup> Enantiopure intermediate 5 as well as derived enone 7 and diene 8 should prove useful in the synthesis of higher sugars and related polyols.<sup>16</sup>

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## **References and Notes**

- Johnson, C. R.; Adams, J. P.; Bis, S. J.; De Jong, R. L.; Golebiowski, A.; Medich, J. R.; Penning, T. D.; Senanayake, C. H.; Steensma, D. H.; Van Zandt, M. C. Pure & Appl. Chem. 1992, 64, 1115.
- (a) Johnson, C. R.; Penning, T. D. J. Am. Chem. Soc. 1988, 110, 4726; (b) Medich, J. R.; Kunnen, K. B.; Johnson, C. R. Tetrahedron Lett. 1987, 28, 4131; (c) Johnson, C. R.; Senanayake, C. H. J. Org. Chem. 1989, 54, 735; (d) Johnson, C. R.; Golebiowski, A.; McGill, T. K.; Steensma, D. H. Tetrahedron Lett. 1991, 32, 2597; (e) Johnson, C. R.; Ple, P. A.; Su, L.; Heeg, M. J.; Adams, J. P. Synlett 1992, 388.
- Johnson, C. R.; Sakaguchi, H. Synlett 1992, 813; Sundram, H.; Golebiowski, A.; Johnson, C. R. Tetrahedron Lett. 1994, 35. (in press).
- For examples of new synthetic methodologies developed in the course of this project see: (a) Johnson, C. R.; Adams, J. P.: Braun, M. P.; Senanayake, C. B.; Wovkulich, P. M.; Uskokovic, M. R. Tetrahedron Lett. 1992, 33, 919; (b) Bakale, R. P.; Johnson, C. R.; Scialdone, M. A. J. Am. Chem. Soc. 1990, 112, 6729; (c) Johnson, C. R.; Golebiowski, A.; Steensma, D, H.; Scialdone, M. A. J. Org. Chem. 1993, 58, 7185; (d) Johnson, C. R.; Raheja, R. K. J. Org. Chem. 1994, 59, 2287.
- (a) Johnson, C. R.; Golebiowski, A.; Steensma, D. H. J. Am. Chem. Soc. 1992, 114, 9414; (b) Johnson, C. R.; Adams, J. P.; Collins, M. A. J. Chem. Soc. Perkin Trans. 1 1993, 1; (c)

Johnson, C. R.; Golebiowski, A.; Braun, M. P.; Sundram, H. *Tetrahedron Lett.* **1994**, *35*, 1833; (d) Johnson, C. R.; Kozak, J. *J. Org. Chem.* **1994**, *59*, 2910.

- 6. This part of the synthesis was done in analogy to the previously reported 1,4-cyclohexadiene functionalization (see ref. 2e).
- All syn oxygen products are dominant in additions of singlet oxygen to 6-silyloxy-1,4cycloheptadienes (see ref. 2c, 2d, 5a). For a recent review on singlet oxygen cycloadditions see: Clennan, E. L. Tetrahedron 1991, 47, 1343.
- 8. For a recent review on the use of lipases in the presence of isopropenyl acetate see Wong, C.-H.; Whitesides, G. M. *Enzymes in Synthetic Organic Chemistry*; Pergamon : Oxford, 1994
- 9. Chiral HPLC: CHIRALCEL OB (J. T. Baker); 40% isopropyl alcohol in hexane at 0.5 mL/min; ent-6 13.6 min; 6, 15.5 min. A racemic sample of 6 was prepared from 4.
- 10. Moore, R. E.; Barchi, J. J.; Bartolini, G. J. Org. Chem. 1985, 50, 374. No polarimetric data is given for 3-deoxy-D-ribo-hexitol.
- 11. Johnson, C.R.; Braun, M. P. J. Am. Chem. Soc. 1993, 115, 11014.
- 12. Gemal, A. L.; Luche, J-L. J. Am. Chem. Soc. 1981, 103, 5454.
- 13. Baillargeon, V. P.; Stille, J. K. J. Am. Chem. Soc. 1983, 105, 7176.
- Saksena, A. K.; Mangiaracina, P. Tetrahedron Lett. 1983, 24, 273. For an excelent review on substrate-directable reactions see: Hoveyda, A. H.; Evans, D. A.; Fu, G. C. Chem. Rev. 1993, 93, 1307.
- 15. Kazlauskas, R. J.; Weissfloch, A. N. E.; Rappaport, A. T.; Cuccia, L. A. J. Org. Chem. 1991, 56, 2656.

## 16. Selected Spectral and Polarimetric Data:

1: mp 99 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ: 4.63-4.51 (m, 2H); 4.40-4.31 (m, 2H); 2.28-1.93 (m, 8H); 1.44 (s, 3H); 1.34 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ: 107.1, 76.6, 58.0, 31.5, 30.0, 27.7, 25.4. 3: IR (film) : 1450, 1431, 1395, 1357, 1200, 1157 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ : 6.39 (dd, 2H,J = 3.7 Hz, 2.15 Hz); 4.80-4.76 (m, 2H); 4.32-4.23 (m, 2H); 2.56-2.51 (m, 4H); 1.47 (s, 3H); 1.28 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ: 130.6, 105.4, 74.7, 73.7, 37.9, 27.4, 23.9. 4: mp 127-128 °C; IR (film) : 3404, 3337, 1645, 1210, 1070, 1024 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>) δ: 5.75 (d, 2H, J = 3 Hz); 4.41-4.36 (m, 4H); 2.38-2.15 (m, 6H); 1.42 (s, 3H); 1.30 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$ : 134.1, 102.1, 76.3, 66.4, 40.3, 27.3, 24.4. 5: Oil;  $|\alpha|_D$  +23.2 (c 1, CHCl<sub>3</sub>); IR (neat) : 3454, 2978, 1730, 1711, 1366, 1241, 1027 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ : 5.76 (ddd, 1H, J = 11.5, 5.5, 1.8 Hz); 5.63 (ddd, 1H, J = 11.5, 5.5, 1.8 Hz); 5.29-5.22 (m, 1H); 4.59-4.48 (m, 2H); 4.40-4.35 (m, 1H); 2.34-2.07 (m, 5H); 2.06 (s, 3H); 1.40 (s, 3H); 1.32 (s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) & 170.3, 135.5, 129.8, 107.7, 76.9, 76.7, 68.7, 66.9, 41.7, 39.0, 27.4, 24.6, 21.1. 6: mp 49-52 °C; ; [α]<sub>D</sub> -14.9 (c 1, CHCl<sub>3</sub>); IR (cast film) : 1733, 1636, 1366, 1229, 1059, 1016 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>) d: 6.12 (ddd, 1H, J = 13.3, 4.8, 1.1 Hz); 5.97 (d, 1H, J = 13.8 Hz); 5.55-5.48 (m, 1H); 4.52 (ddd, 1H, v = 13.8 Hz); 5.55-5.48 (m, 200); 10.55 (ddd, 1H, v = 13.8 Hz); 10.55 (ddd, 1Hz); 10.55 (10.8, 5.3, 3.3 Hz); 4.35 (ddd, 1H, J = 10.7, 5.2, 3.7 Hz); 2.98 (dd, 1H, J = 17.7, 10.8 Hz); 2.73 (dd, 1H, J = 18.2, 3.4 Hz); 2.34 (ddt, 1H, J = 14.6, 3.7, 1.4 Hz); 2.07 (s, 3H); 1.43 (s, 3H); 1.35(s, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>) δ: 201.7, 169.6, 134.9, 131.4, 107.8, 74.5, 73.1, 67.9, 44.4, 32.0, 27.7, 25.5, 20.7. 9 (from cyclooctadiene 8):  $[\alpha]_D$  +0.4,  $[\alpha]_{400}$  +1.6 (c1, CHCl<sub>3</sub>); 9 (from cyclopentene 12):  $[\alpha]_D$  +0.4 (c 3.6, CHCl<sub>3</sub>),  $[\alpha]_{4(0)}$  +1.8 (c2.1, CHCl<sub>3</sub>); The following spectral data were identical in both cases: IR (neat) : 1742, 1433, 1370, 1235, 1045, 948 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$ : 5.21-5.28 (m, 3H); 4.28 (dd, 1H, J =4.7, 3.7 Hz); 4.24 (dd, 1H, J = 4.7, 3.7 Hz); 4.12 (dd, 1H, J = 12.2, 6.1 Hz); 4.03 (dd, 1H, J = 12.2, 6.1 Hz); 2.04 - 2.07 (m, 17H);  $^{13}$ C NMR  $(CDCl_3)$   $\delta$ : 170.5, 170.0, 170.2, 71.5, 68.4, 68.3, 64.3, 61.6, 31.2, 20.9, 20.8, 20.6

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